

**Working with Broadband Transient Absorption Data in Igor Pro**  
**v. 4.4 November 29, 2012**

**Table of Contents**

<b>I. VERSION AND EXTENSION REQUIREMENTS</b>	<b>1</b>
VERSION REQUIREMENTS	1
LOADING THE MULTipeakFit XOP	1
<b>II. IMPORTING TRANSIENT ABSORPTION DATA INTO IGOR PRO</b>	<b>2</b>
DATA FILE FORMAT REQUIREMENTS	2
LOADING DATA FILES INTO IGOR	2
WAVE FORMAT	5
<b>III. PLOTTING TA DATA</b>	<b>7</b>
AUTOMATICALLY GENERATED PLOTS	7
3D SURFACE PLOTS	8
SCATTER AND CONTOUR PLOTS	9
IMAGE PLOTS	9
<b>IV. VISUALIZING SPECTRA AND KINETICS FROM AN IMAGE PLOT</b>	<b>11</b>
QUICK VISUALIZATION USING THE DATA INSPECTOR	11
EXTRACTING 1D DATA FOR FITTING AND PLOTTING	12
<b>V. CURVE AND PEAK FITTING</b>	<b>14</b>
<b>VI. DATA PROCESSING PROCEDURES</b>	<b>16</b>
TIME ZERO CORRECTION	16
SCATTERED LIGHT CORRECTION	17
EXTRACT REGION OF INTEREST	20
COMBINE DATA SETS	22
<b>VII. CHIRP CORRECTION PROCEDURES</b>	<b>24</b>
IMPORT FIT COEFFICIENTS GENERATED WITH SX PRO	24
GENERATE FIT COEFFS FROM SOLVENT RESPONSE	24
GENERATE T0 WAVE FROM FIT COEFFS	25
CHIRP CORRECTION FROM T0 WAVE	26
<b>VIII. DATA ANALYSIS USING SINGULAR VALUE DECOMPOSITION</b>	<b>28</b>
INTRODUCTION	28
SINGULAR VALUE DECOMPOSITION	28
RANK SVD MATRICES	30
GLOBAL FIT	31
SPECIES ABSORPTION FROM GLOBAL FIT	31
<b>IX. EXPORT A PROGRESSIVE TIME AXIS FOR DATA ACQUISITION</b>	<b>32</b>

<b>APPENDIX A: AUTOMATICALLY GENERATED WAVE NAMES</b>	<b>34</b>
---	-----------

<b>APPENDIX B: DESCRIPTION OF FITTING FUNCTIONS AND COEFFICIENTS</b>	<b>36</b>
--	-----------

*Please direct questions and comments to the author:*

Matt Sfeir

[msfeir@bnl.gov](mailto:msfeir@bnl.gov)

## I. Version and Extension Requirements

### Version Requirements

These procedures require Igor Pro version 6.2 or later. Igor Pro is produced by Wavemetrics, Inc. More information can be found at:

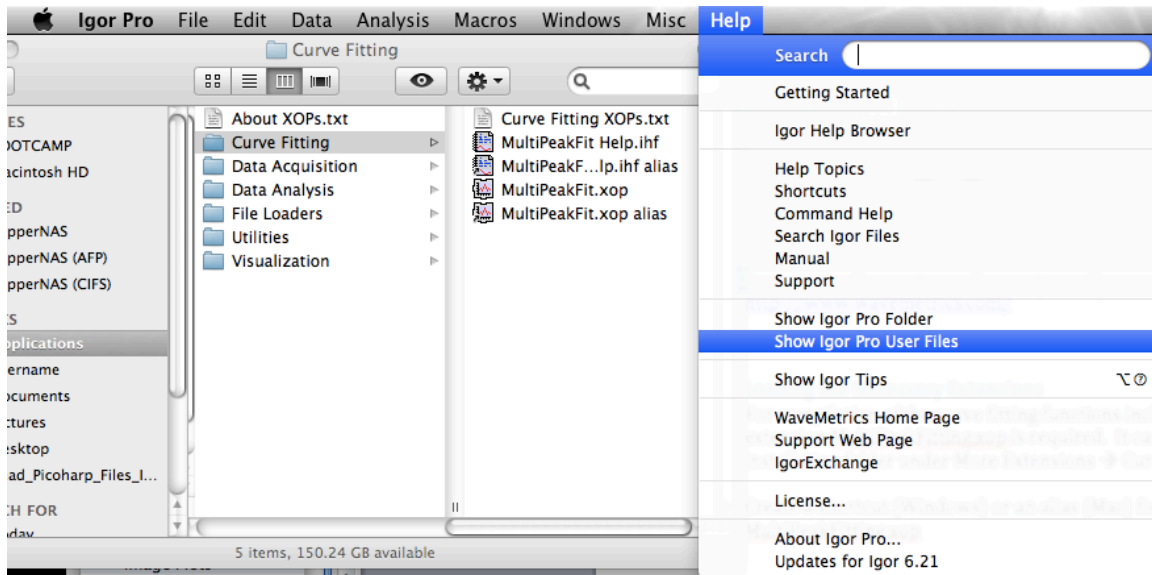
<http://www.wavemetrics.com/>

### Loading the MultiPeakFit XOP

Included in the Igor installation, but not loaded by default is the MultiPeakFitting.xop extension. This file is required for data analysis and must be loaded before the procedures are compiled, particularly for the curve fitting functions. It can be found in the Igor Pro installation folder under More Extensions → Curve Fitting → MultiPeakFitting.xop

*Note: Compilation of these procedures will fail unless this extension is loaded!*

To set up your Igor Pro environment to load this file by default at startup:  
Create a shortcut (Windows) or an alias (Mac) for the files *MultiPeakFit.xop* and *MultiPeakFitHelp.ihf*.



From the Igor Help menu, choose, “Show Igor Pro User Files.” Add these shortcuts to the Extensions folder in the User files directory.

If Igor is already running, then it must be restarted for these changes to take effect.

## II. Importing Transient Absorption Data into Igor Pro

### Data File Format Requirements

This package includes a set of functions for processing and graphing wavelength and time-resolved absorption data. This code was designed to process data created by the Helios Femtosecond Transient Absorption spectrometer produced by Ultrafast Systems, LLC. However, it will work with any data with the format:

```
0, time_1, time_2, ..., time_M  
wavelength_1, deltaOD_1_1, deltaOD_1_2, ..., deltaOD_1_M  
wavelength_2, deltaOD_2_1, deltaOD_2_2, ..., deltaOD_2_M  
.  
.  
.  
wavelength_N, deltaOD_N_1, deltaOD_N_2, ..., deltaOD_N_M
```

where  $N, M$  are arbitrary integers.

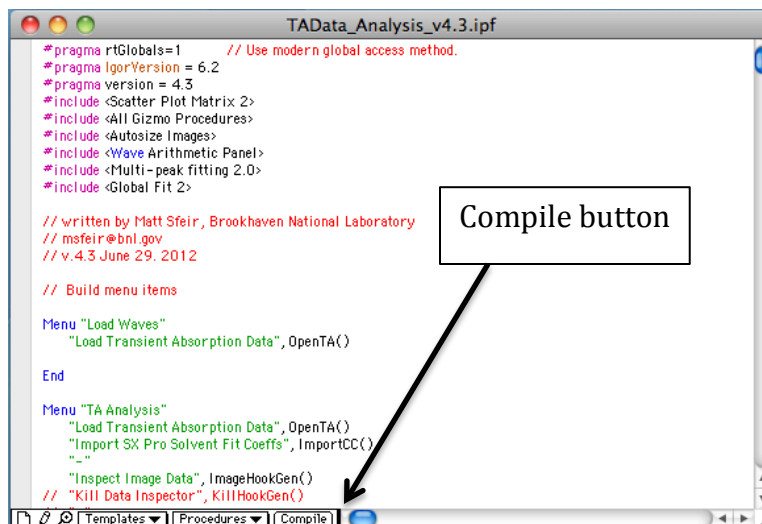
### Loading Data Files into Igor

- 1) Load and compile the appropriate procedures into a new Igor Pro experiment, using the appropriate menu options.

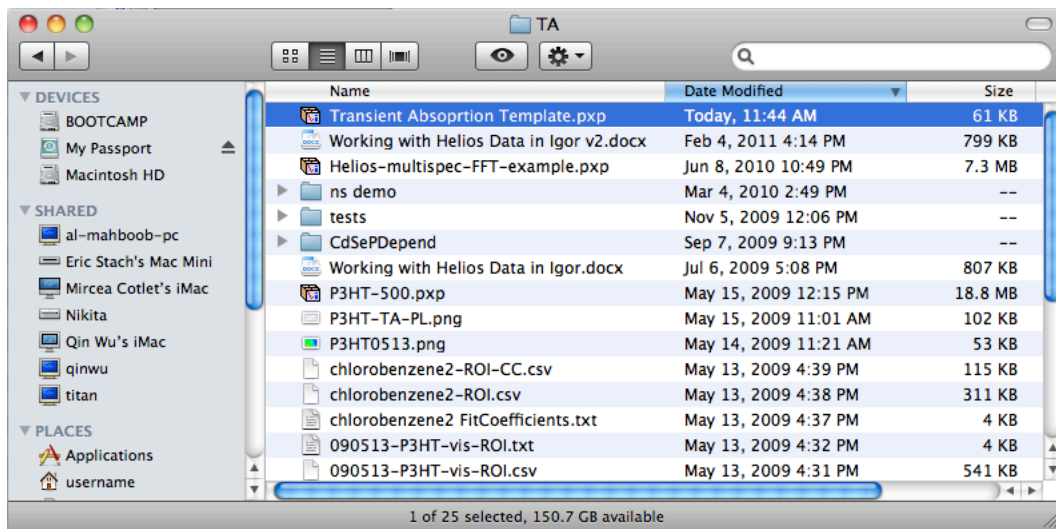
File → Open File → Procedure ...

*TADData\_Analysis\_vX.XX.ipf*

*TADData\_CurveFitting\_vX.XX.ipf*

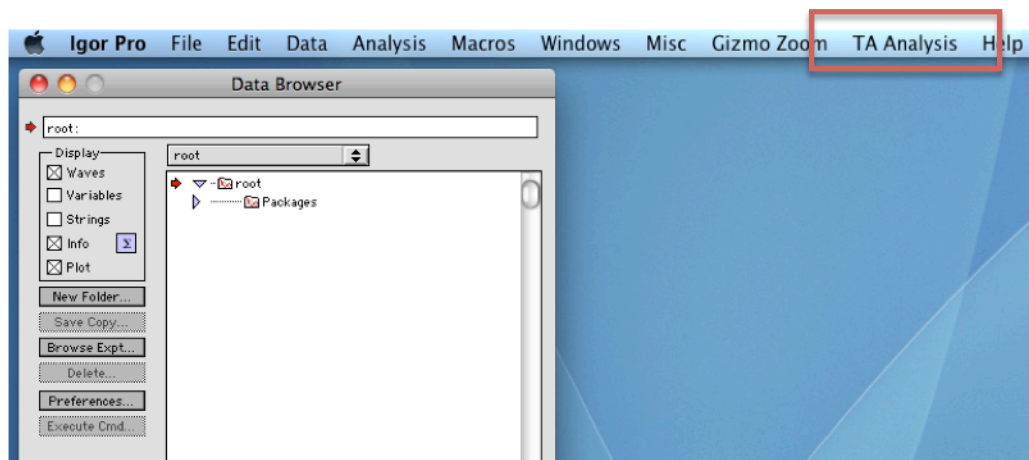


- 2) Alternatively, open the Igor Packed Experiment file named “Transient Absorption Template.pxp”. Note that this makes upgrading to a new procedure file more difficult.

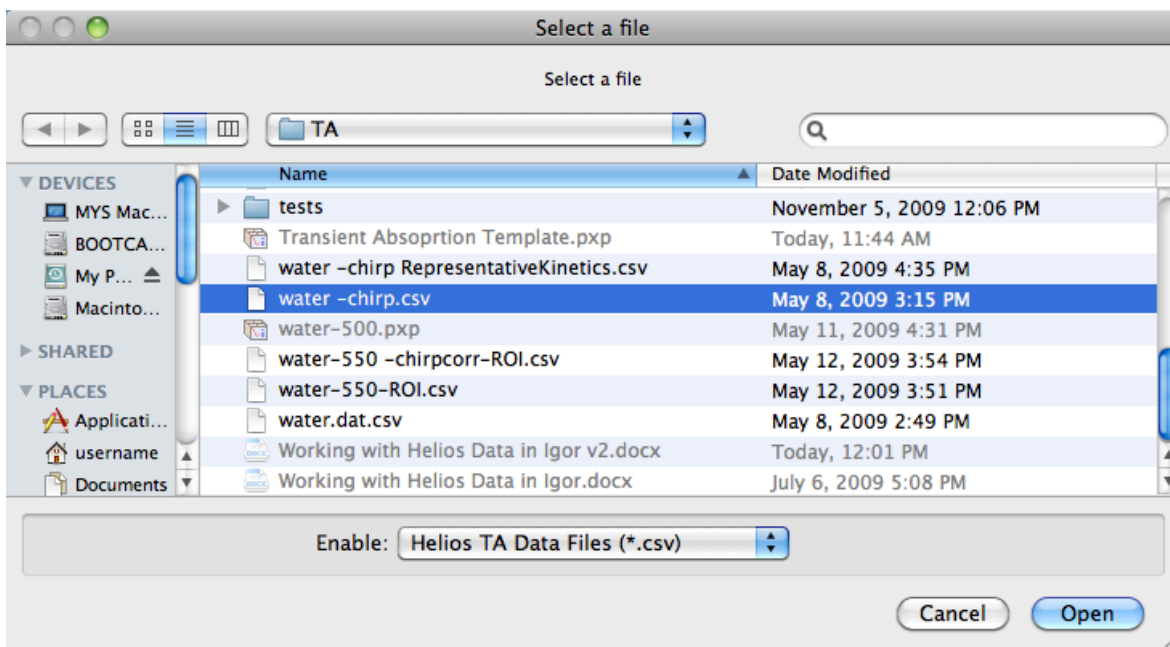
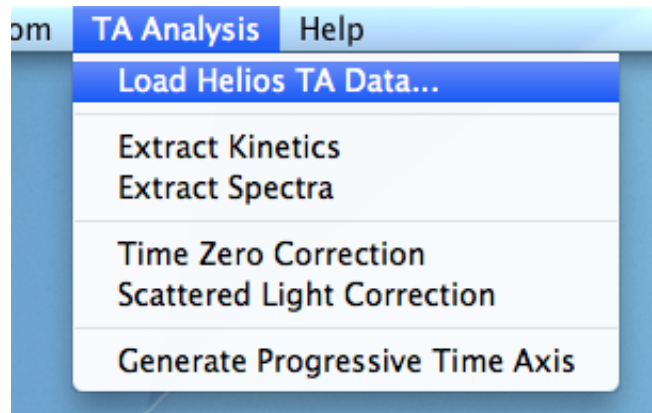


If the template is launched, a blank experiment will open which automatically includes and compiles the necessary procedures.

- 3) When the procedures are properly loaded, a new menu will appear on the task bar entitled, “TA Analysis.”



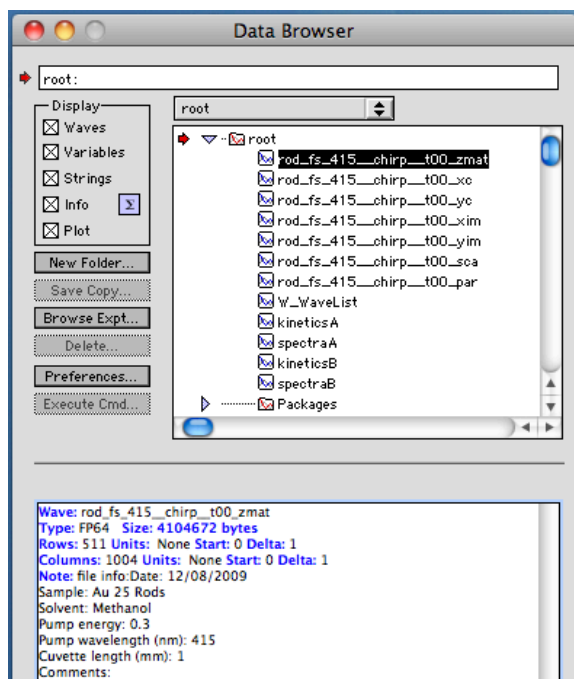
- 4) Open your saved transient absorption data (.csv files) using the appropriate menu option:  
TA Analysis → Load Helios TA Data ...



## Wave Format

- 5) This procedure loads the original data in *filename* and creates a new set of processed waves (*filename\_zmat*; *filename\_xc*; *filename\_yc*). The procedure replaces all the NaNs and Infs in the original file with 0 and deletes extra rows (for which the spacing is non-monotonic). It also creates several additional waves for graphing purposes:

*filename\_zmat* (corrected matrix)  
*filename\_xc* (processed row values)  
*filename\_yc* (processed column values)  
*filename\_xim* (x values for image plot)  
*filename\_yim* (y values for image plot)  
*filename\_sca* (xyz wave for scatter plot)  
*filename\_par* (3D parametric wave)



The footer information in the original file (beginning with “file info”) is stored in *filename\_zmat* as an Igor Note. This information can be viewed directly from the Data Browser by highlighting the appropriate wave. Alternatively, it viewed from any plot, by holding down Command-Option-Control (Mac) or SHIFT+F1 (Windows) and clicking on the graph.

By default, the Image Inspector window hook is enabled for imported data files. This allows you to graphically inspect the kinetic and spectral information contained in a given data set. The waves, *kineticsA*, *spectraA*,

*kineticsB*, *spectraB*, and *W\_Wavelist* are used by this procedure and can be ignored.

*Note: Starting in version 4.1, the procedure will now remove the original, unprocessed data file and associated row and column position waves including: RP\_filename (wavelength labels for row data in nm); filename (original uncorrected matrix in  $\Delta OD$ ); CP\_filename (time labels for column data in ps).*

*These can be reintroduced by commenting out the following line in the ProcessTA() function:*

`KillWaves $wName, $RPName, $CPName`

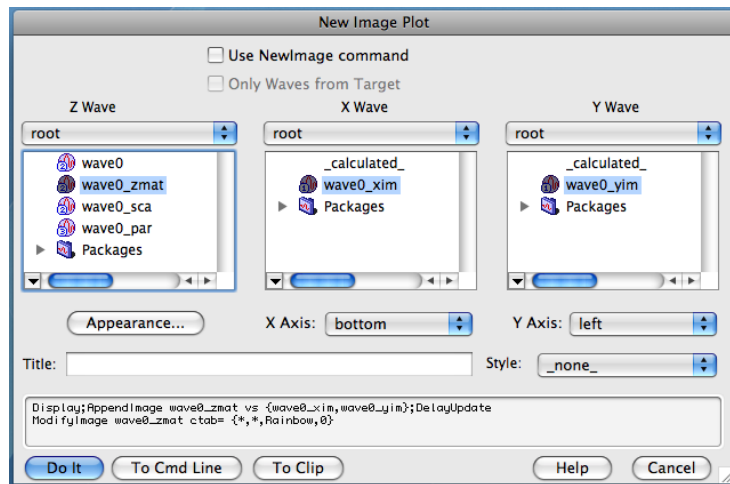


### III. Plotting TA Data

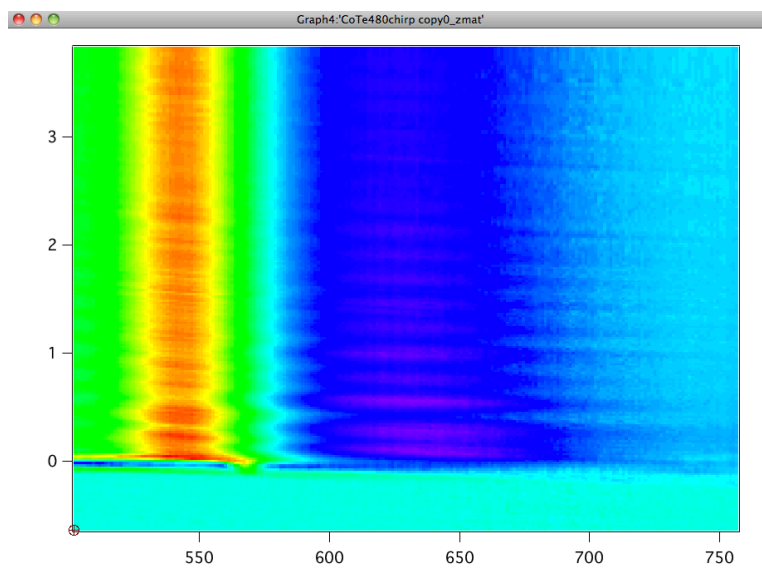
#### Automatically Generated Plots

- 1) An image plot of your data will be automatically created with a visible cursor.  
This plot can be generated manually using *filename\_zmat*, *filename\_xim*, and *filename\_yim*:

Windows → New → Image Plot



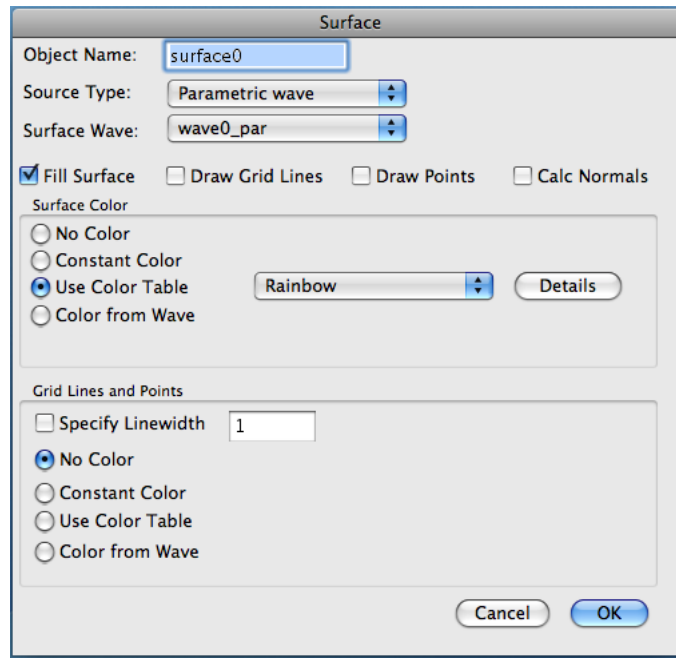
Click the “Appearance” button to generate a color image.



Use the CTRL-I (CMD-I for Mac) key sequence to manually show the cursors.

### 3D Surface Plots

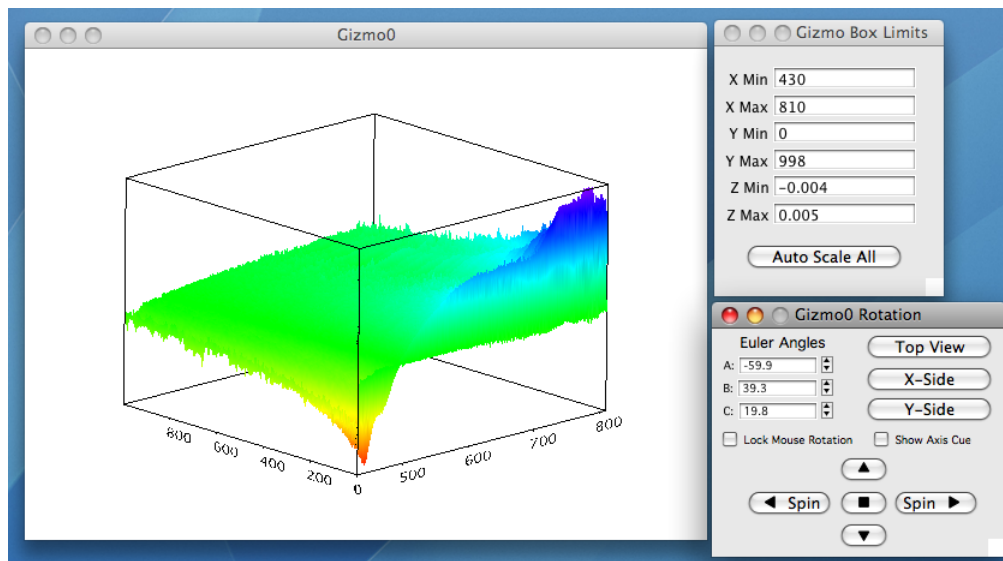
- 2) To generate a 3D Plot of your data:  
Windows -> New -> 3D Plots -> Surface Plot



Modify as appropriate:

Gizmo -> Axis Range

Gizmo -> Rotation

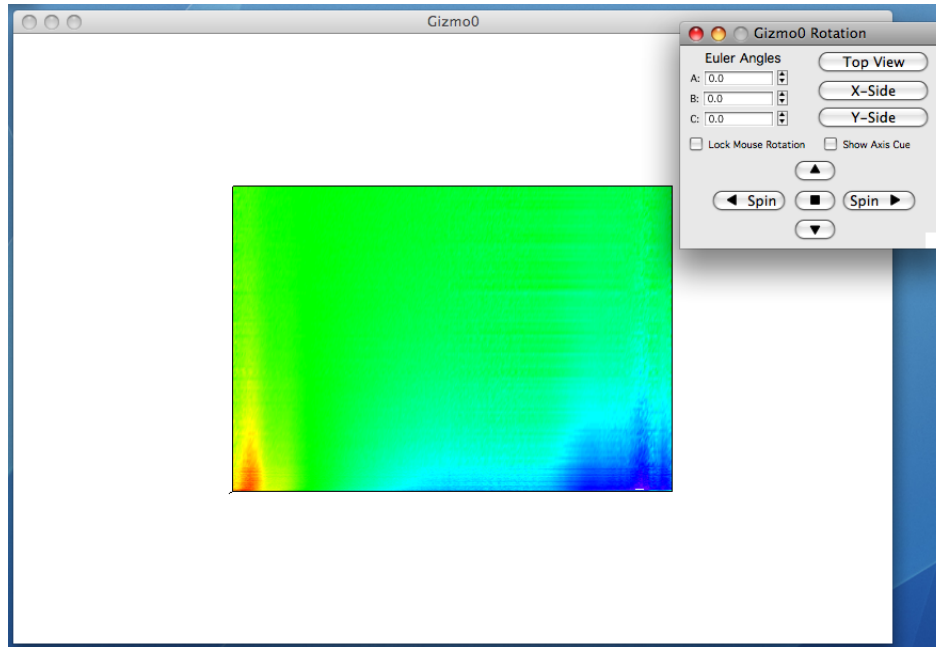


## Scatter and Contour Plots

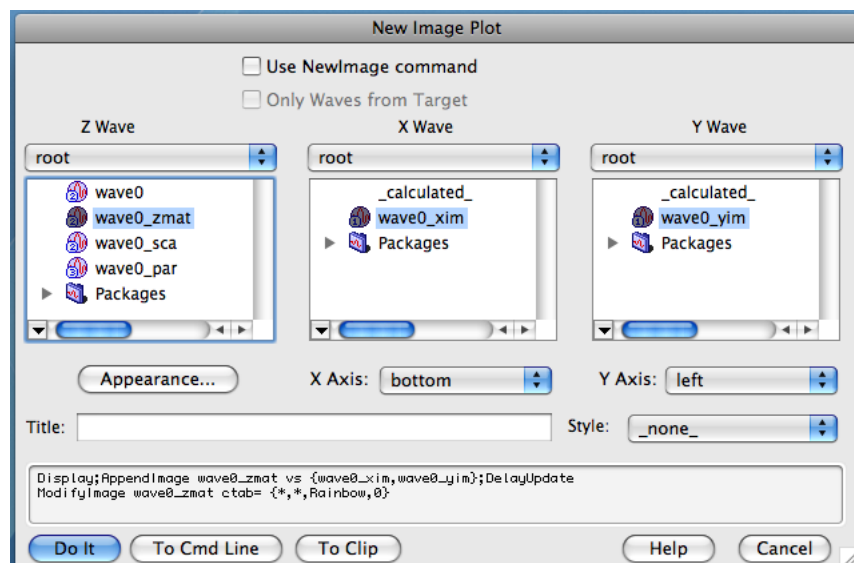
- 3) Similar procedures can be used to create 3D scatter plots and 2D contour plots.

## Image Plots

- 4) Image Plots can be generated in one of the following ways:
  - A. In the Gizmo rotation panel, click "Top View."



- B. The ProcessTA procedure generates axes which are *approximately* scaled to generate an image ( $N \rightarrow N+1$  points with no interpolation).  
Windows  $\rightarrow$  New  $\rightarrow$  Image Plot

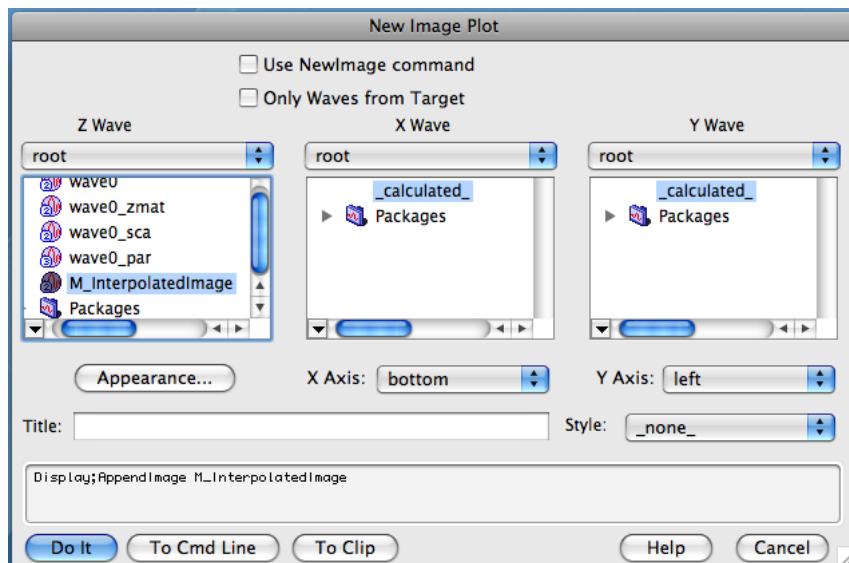


Click the “Appearance” button to generate a color image.

C. Interpolate a particular region of interest. Image interpolation can take several minutes depending on the image size and the mesh. An example command would be:

`ImageInterpolate /S={600,1,800,0,0.1,10} Voronoi wave0_sca`

This will generate an interpolated image over the spectral range 600 – 800 nm (1 nm mesh) and time range from 0 – 10 ps (100 fs mesh). On a 2.5 GHz Intel Core 2 duo processor with 4 GB of RAM, this command takes ~ 9 minutes on an original XYZ data set that contains 109056 rows.

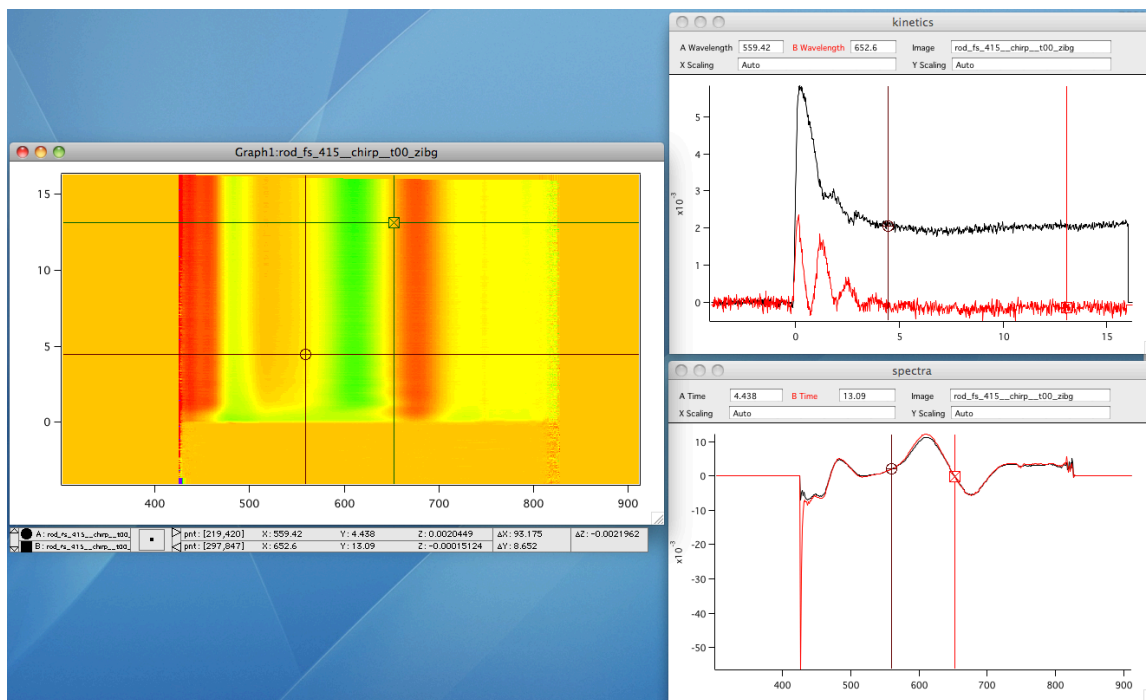


## IV. Visualizing Spectra and Kinetics from an Image Plot

### Quick Visualization Using the Data Inspector

Spectra or kinetic data can be visualized through a set of windows that are automatically created when an image file is loaded or generated via one of the built-in processing functions.

The windows are named “kinetics” (vertical slices) and “spectra” (horizontal slices). Cursors A and B can be used to look at your data at a fixed point in time or wavelength.

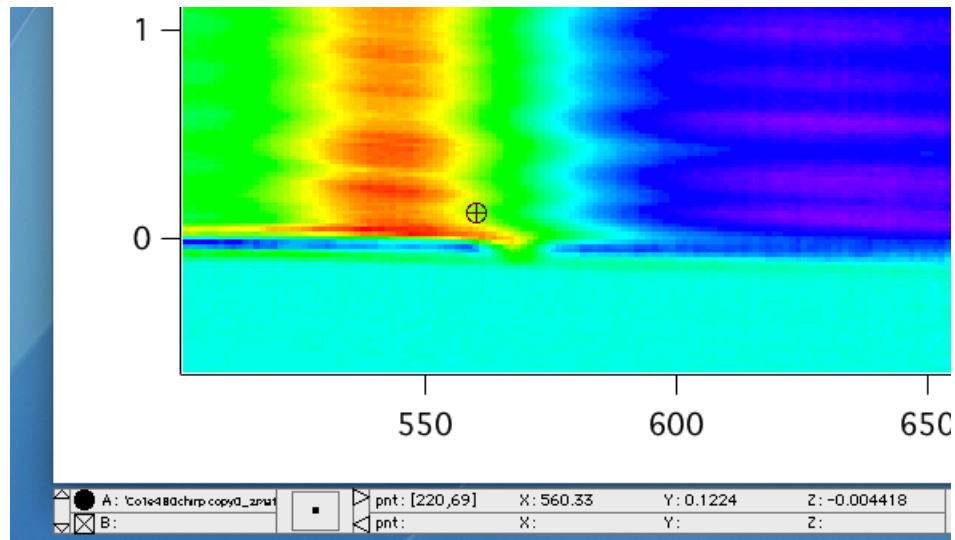


The information displayed in these windows is automatically updated when the cursors are moved or when a different image plot is activated.

Data in these windows should not be fit or manipulated directly. Use the “Extract ...” functions described in the next section to save regions of interest.

## Extracting 1D Data for Fitting and Plotting

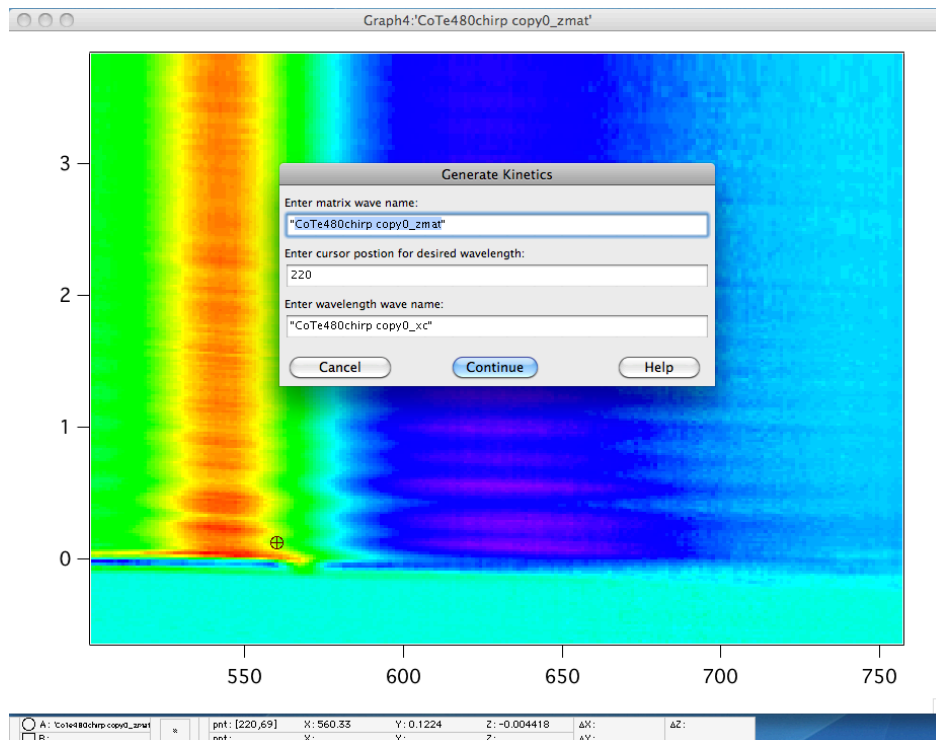
- 1) Extract Spectra and Kinetics from an image from *filename\_zmat*.  
The simplest way is to use the automatically generated image plot. Place the cursor in the spot where the data is to be extracted:



Select the appropriate function from the TA analysis menu:

TA Analysis → Extract Kinetics

TA Analysis → Extract Spectra



The procedure will attempt to auto-fill these values for you:

Matrix wave name: filename\_zmat

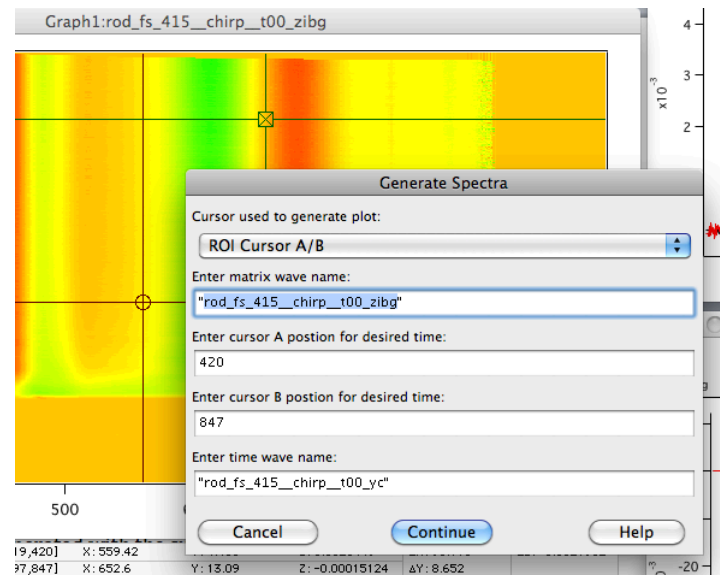
Cursor position: element n [m] for (n,m) matrix for kinetics [spectra]

- note: this is not the actual wavelength or time value

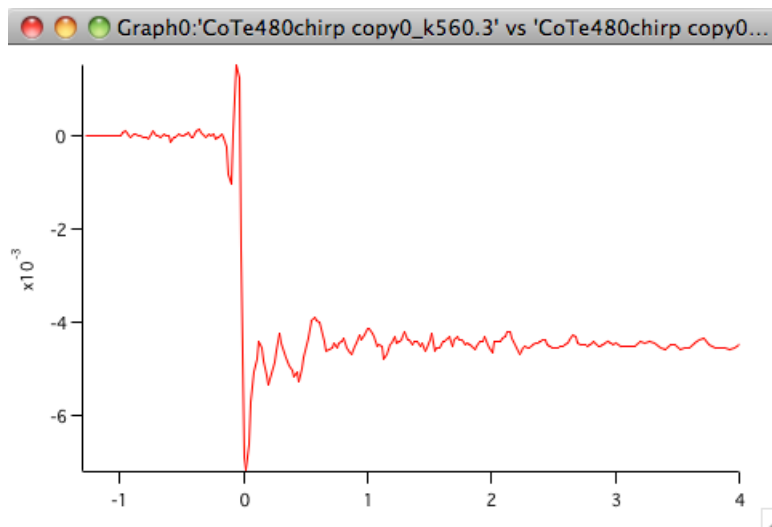
Wavelength [time] wave name: typically filename\_xc [filename\_yc]

- note: this is not the waves used generate the image plot (which contain N+1 points rather than the desired N)

If both Cursors A and B are on the graph, you will have the option of extracting the data at either position or averaging between them:

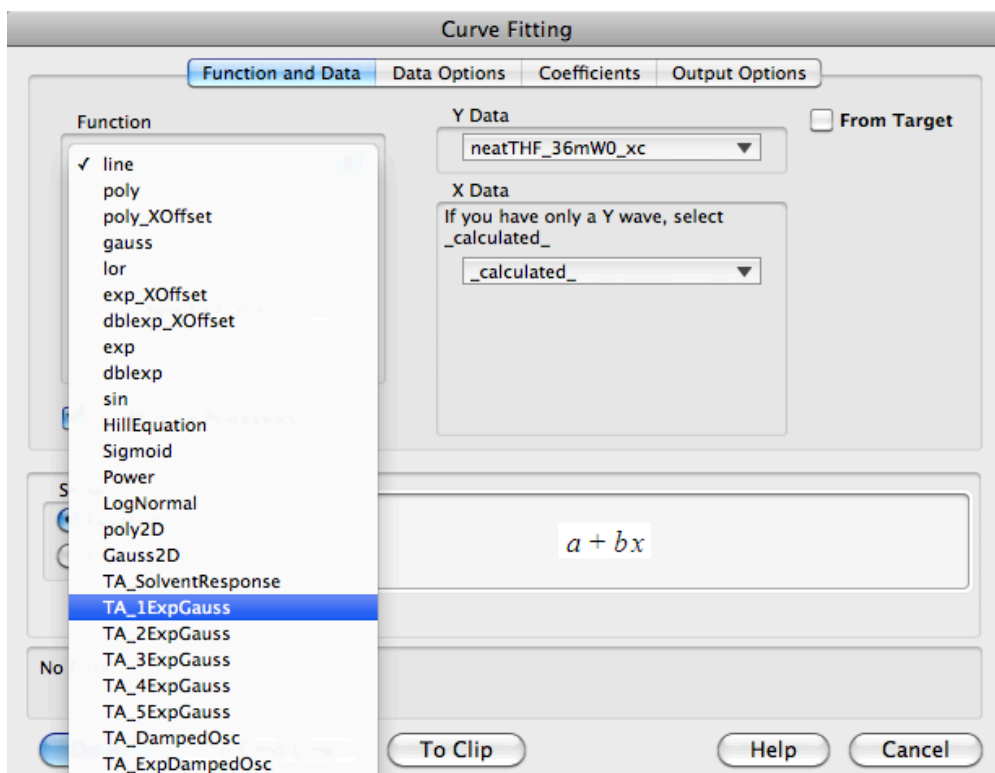


New waves will be generated with the suffix *\_kwavelength\_pix* or *stime\_pix* (pix: number of pixels in the average of A and B cursors). 1D plots will be generated automatically if the expected file structure exists.



## V. Curve and Peak Fitting

- 1) Fits of the kinetic data can be made using the built in Curve Fitting functions.  
Analysis → Curve Fitting →



A few special fitting functions are added in this experiment:

TA\_SolventResponse

TA\_nExpGauss

TA\_nExpGaussDampedOsc

These use the built-in ExpGauss function (from the MultiPeakFit extension) to convolute the effect of impulsive excitation and exponential decay. The fit functions are constructed so that a common pulsewidth and common time zero is used for all exponentials.



For examples, the parameters for TA\_nExpGauss are described below. Initial guessss must be entered in the Coefficients tab on the Curve Fitting Dialog window:

y0: constant y offset, usually fixed at 0 during fitting

pw: pulsewidth can be usually set to .12 for initial guess.

- Note: This is the standard deviation of the Gaussian used to approximate the laser pulse, not the FWHM. The conversion is  $\text{FWHM} \sim 2.355 * \text{pw}$ .

t0: time zero (time of impulsive excitation)

ampn: amplitude of first exponential decay

Gn: the rate of the first exponential decay component

- Note: This is equal to the inverse of the lifetime

For a detailed description of fitting functions and their use, please refer to Appendix B. This information will be updated as necessary.

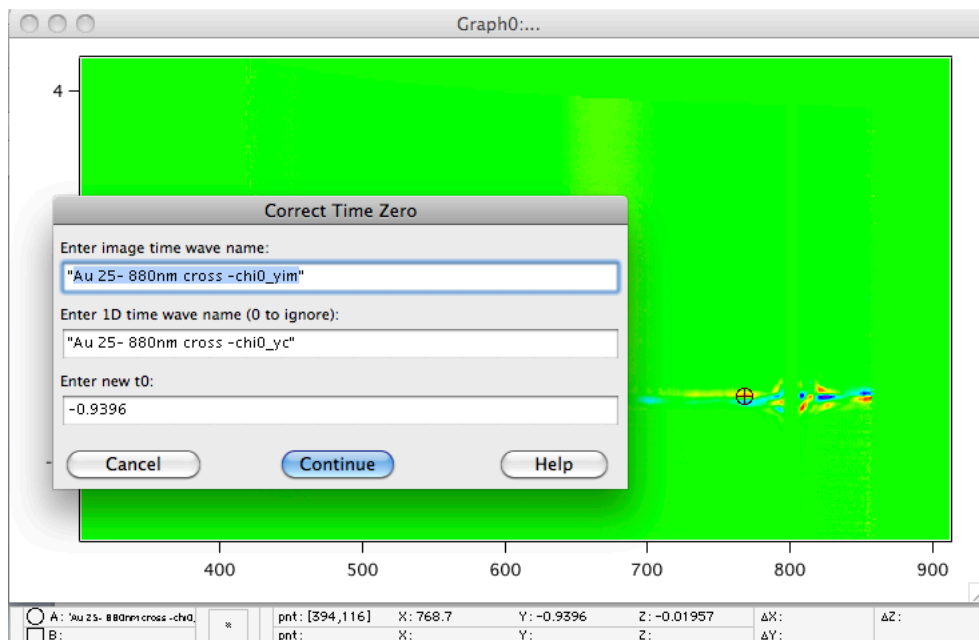
- 2) Transient spectra can be fit using the Multi-peak fit dialog.  
Analysis → Multi-peak Fit → Start New Multi-peak Fit

More information can be found in the Igor Help Browser.

## VI. Data Processing Procedures

### Time Zero Correction

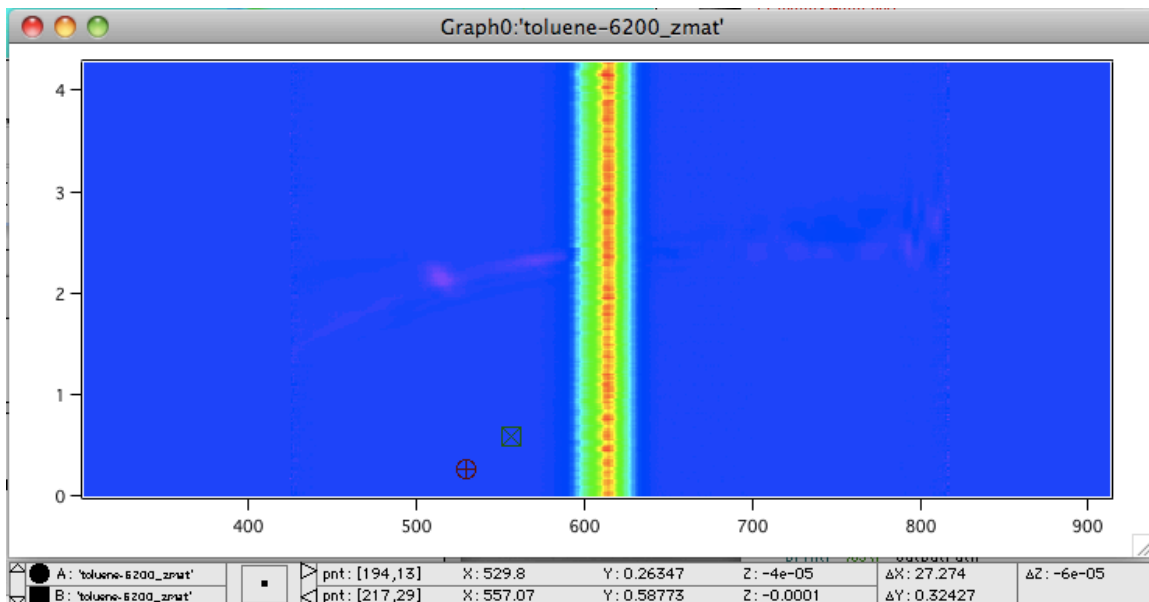
- 1) Use this procedure to correct your image and spectra for an absolute time zero. Place cursor A on your image at time zero and choose:  
TA Analysis → Time Zero Correction
- 2) This procedure attempts to automatically pick the correct time files and will choose the cursor position as the new time zero. These can be modified from the dialog prompt:



- 3) The first wave name, *filename\_yim* is the y axis for the image plot (N+1 point) and the second, *filename\_yc*, is the y axis for extracted kinetics (N points). To leave the kinetic plots undisturbed, enter "0" in this field. To choose a different value for the new time zero, manually enter it into the appropriate field.

## Scattered Light Correction

- 1) Scattered light collected by the array detectors can be automatically removed using the “Scattered Light Correction” procedure. The easiest way to do this is to use an image plot of your data.
- 2) Place cursor A and cursor B on the image to select a range to be averaged for the correction. Both of these cursors should be in the  $t < 0$  regime. Avoid early times that contain empty pixels (due to chirp correction).



- 3) Choose the appropriate procedure from the TA Analysis menu:  
TA Analysis → Scattered Light Correction

Get Cursor Positions

Scattered Light Correction

Cursor A: 67    Cursor B: 43

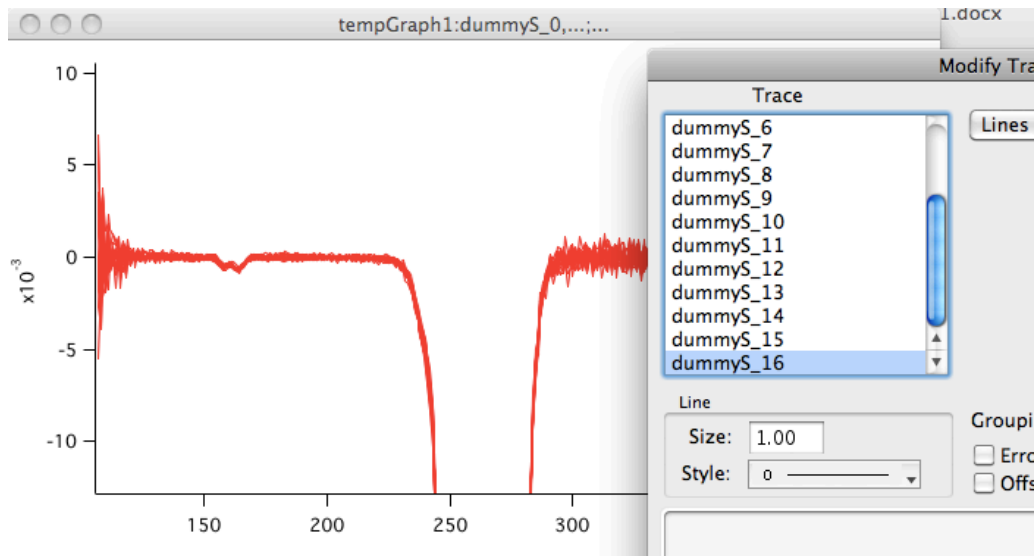
Preview    Save

matrix: toluene-6200\_zmat

sca\_bkgd: toluene-6200\_scarb

corrected: toluene-6200\_zibg

- 4) Click the preview button to view the set of curves that will be averaged to make the correction file. The locations of the cursors will be sent automatically, as will the wave names.



- 5) If this range is acceptable, close the window to return to the correction dialog box and click "Save." Make sure that the image file is the top graph, i.e., click on it again to bring it to the front before clicking "Save."

Alternatively, these procedures can be run from the command line interface.

The syntax is:

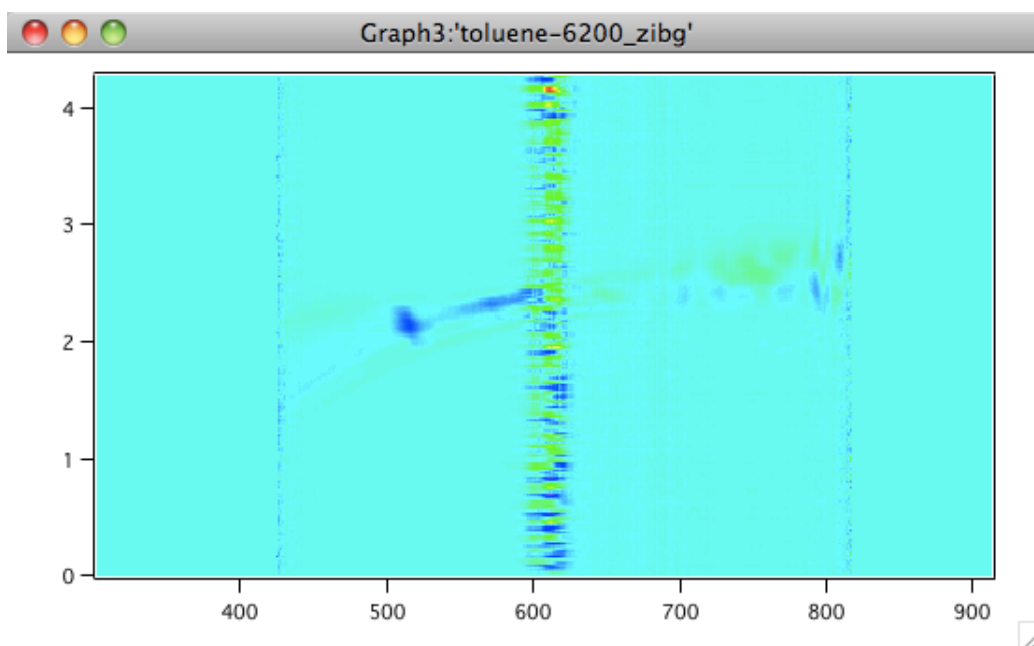
PreviewSLC(filename\_zmat, imin, imax) for Preview

ApplySLC(filename\_zmat, imin, imax) to apply SLC

where *imin* and *imax* are the indices of the time positions to be averaged.

- 6) A new matrix is produced with the wave name *filename\_zibg* along with a copy of the averaged  $t < 0$  file used for the correction (*filename\_scab*).

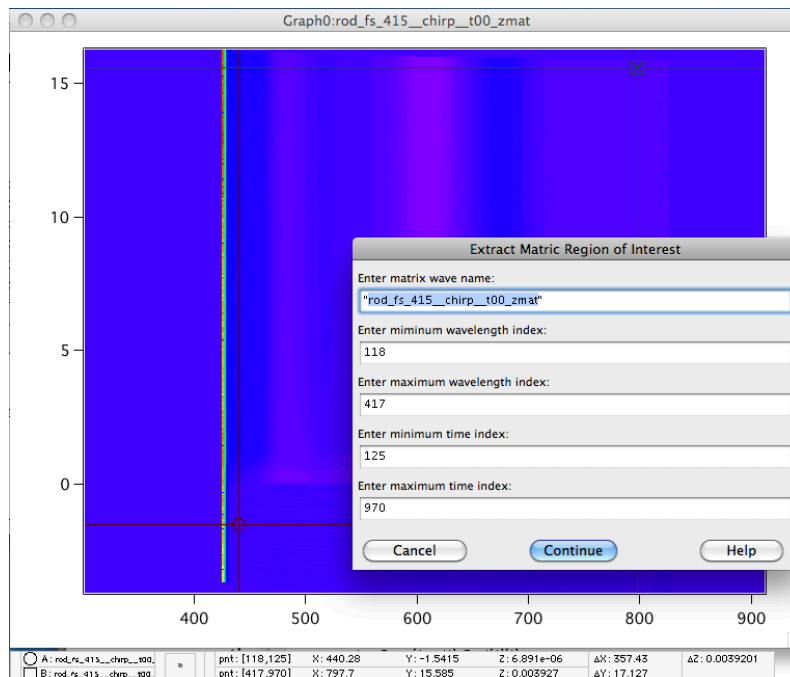
A new image plot is produced with the corrected wave file. The “Extract Spectra” and “Extract Kinetics” routines will also work with the new image file.



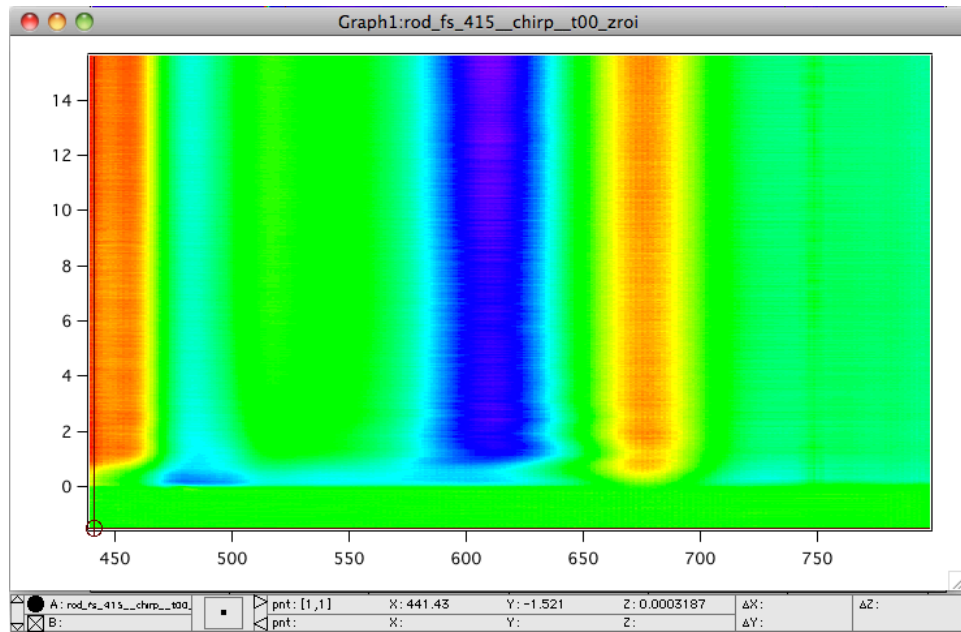
## Extract Region of Interest

For some visualization and analysis purposes, including for Singular Value Decomposition, only a subset of the full dataset is required. For convenience, this function allows you to extract a subset of the data matrix using an image plot and the cursors.

- 1) Place cursor A and B on your image plot such that they define the boundary of the new data set. Click on the image plot to make it the top graph.
- 2) Choose the appropriate procedure from the TA Analysis menu:  
TA Analysis → Extract Region of Interest



- 3) The procedure will attempt to automatically generate the matrix wave name, and matrix indices from the image plot. If the values are acceptable, choose *Continue*.
- 4) A new set of waves will be generated along with an new image plot:
  - filename\_zroi* (subset of zmat)
  - filename\_xroi* (subset of xc)
  - filename\_yroi* (subset of yc)
  - filename\_ximr* (new image x coordinates)
  - filename\_yimr* (new image y coordinates)



- 5) The new set of waves can be operated on by other procedures in the same way as the original *filename\_zmat* wave.

## Combine Data Sets

This function allows you to generate one matrix (for visualization and analysis) from two different data sets that share either an identical time or identical wavelength axis. This can be useful, for example, for combining long and short scans provided that the data sets either starts at the same point on an absolute time scale or are already carefully corrected for time zero.

It will generate a matrix with dimensions equivalent to the number of unique points in the non-identical axis. Points that correspond to identical time (or wavelength) will be averaged and the extra rows are removed so that an image plot can be generated with custom x and y points.

- 1) To execute this function, choose:  
TA Analysis → Combine Data Sets
- 2) Input the wavelength (x), time (y), and  $\Delta A$  values for both sets of data to be combined.

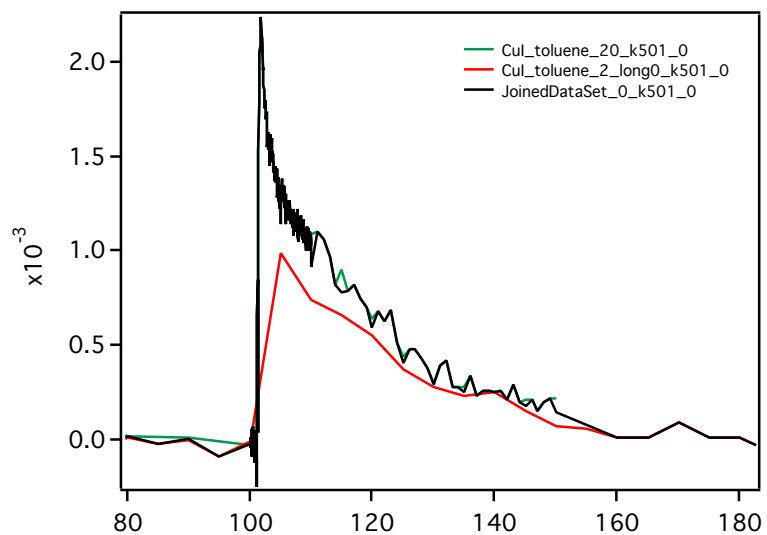
Select data sets to join and sort

Enter X wave for data set 1:	Enter Y wave for data set 1:
<input type="text" value="CuI_toluene_20_xc"/>	<input type="text" value="CuI_toluene_20_yc"/>
Enter Z matrix for data set 1:	Enter normalization factor for data
<input type="text" value="CuI_toluene_20_zmat"/>	<input type="text" value="1"/>
Enter X wave for data set 2:	Enter Y wave for data set 2:
<input type="text" value="CuI_toluene_2_long0_xc"/>	<input type="text" value="CuI_toluene_2_long0_yc"/>
Enter Z matrix for data set 2:	Enter normalization factor for data
<input type="text" value="CuI_toluene_2_long0_..."/>	<input type="text" value="1"/>

- 3) A scaling factor can be applied to either data set if needed.
- 4) Click continue. A new data set will be generated with the default name, "JoinedDataSet\_n\_," with the appropriate suffix (*xc*, *yc*, *zmat*, *xim*, *zim*) corresponding to the different wave types. These waves are now be operated on with any of the other functions.



5) As an example, here are the kinetic traces for the two original and combined data sets:



## VII. Chirp Correction Procedures

### Import Fit Coefficients Generated with SX Pro

If you have already generated a set of Fit Coefficients from solvent data, then these can be easily imported and used in Igor. Choose:

TA Analysis → Import SX Pro Solvent Fit Coeffs

A new 2D matrix, *filename\_sxfcn* will be generated with the lambda values in the first column and the corresponding time zero points in the second column. Extra information contained in this file, such as amplitude coefficients, is discarded. This file can then be used to correct subsequent data sets using the “Generate t0 wave from Fit Coefficients” and “Chirp Correction from t0 Wave” procedures.

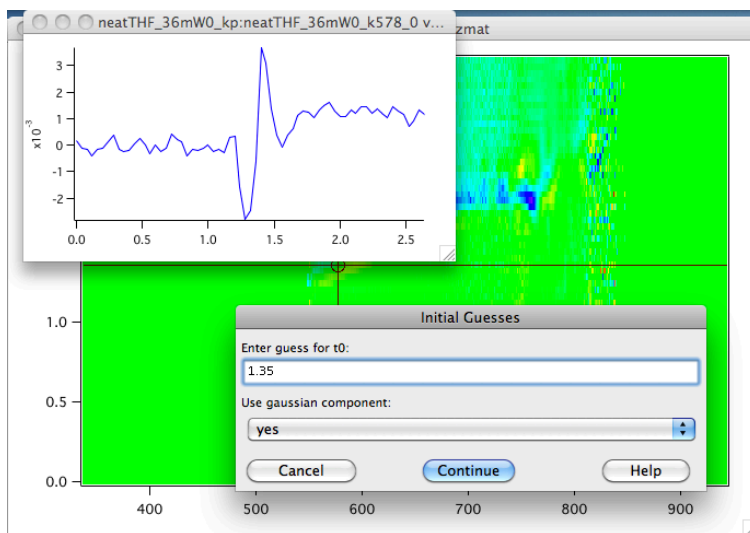
### Generate Fit Coeffs from Solvent Response

This procedure attempts to simplify chirp correction by automatically extracting, fitting, and storing kinetic data using the “SolventResponse” fit function. It will generate a 2D matrix with sets of lambda, time zero pairs in the first and second columns. Alternatively, a similar file can be generated by hand (using and fit function) and used with subsequent chirp correction procedures.

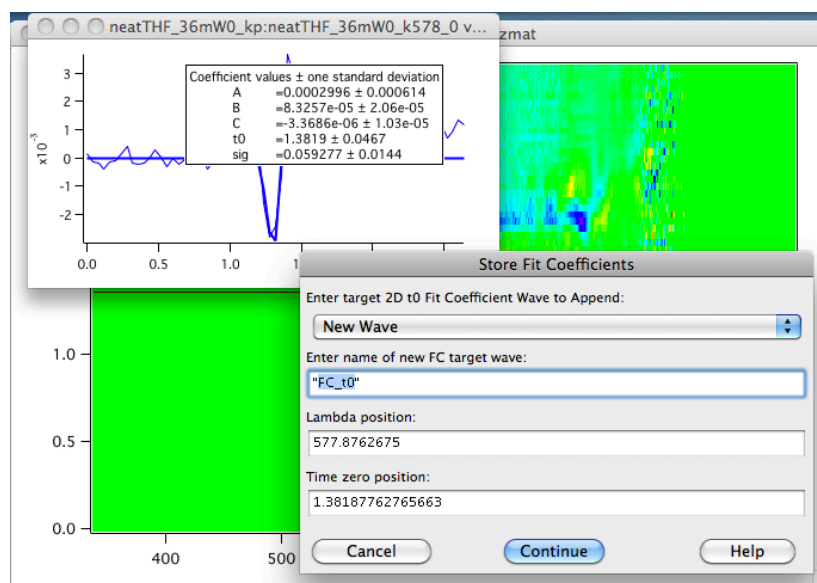
Click on the target image window and position one of the cursors at the wavelength to be fit. Choose:

TA Analysis → Generate Fit Coeffs from Solvent Response

This procedure will first run the “Generate Kinetic” procedure with the standard dialog for matrix wave name, cursor position, and wavelength wave name. See section IV for more details.



Next, a dialog will ask for an initial guess for time zero. I have found that the fit is most sensitive to this value, so be careful with this input. The other initial guesses for amplitudes and pulse width and automatically generated. If you find that this gives you poor fits, then you should generate the appropriate initial guesses using the Curve Fitting Dialog. By default, the fitting procedure uses first and second derivatives of a Gaussian to fit the coherent response of the solvent. If an additional Gaussian component is appropriate, choose “yes” for this option in the dialog box.

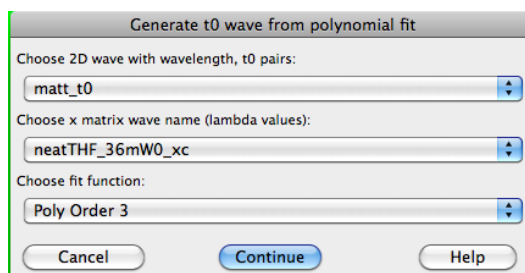


If a good fit is obtained, then click continue to store the lambda and time zero values in an appropriate matrix. Choosing “New Wave” lets you write values to a new matrix. Otherwise choose the appropriate wave so that values may be appended.

This should be repeated with as many points as possible which yield an accurate time zero determination, typically 5 or more.

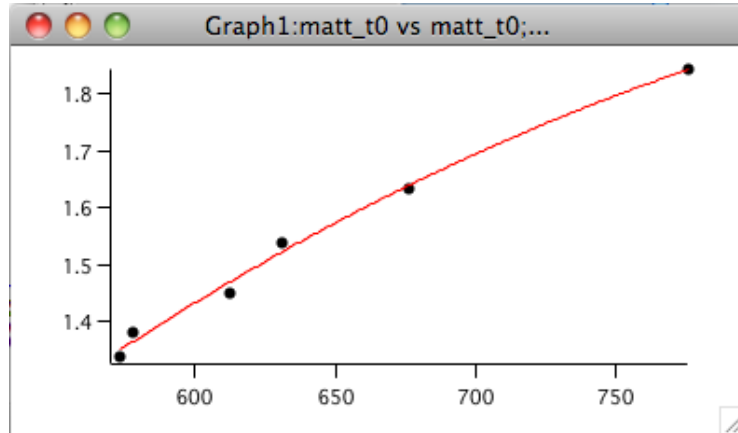
### Generate t0 wave from Fit Coeffs

Once a matrix of lambda, time zero pairs are generated (either manually or using one of the above methods), then they can be used to correct transient absorption data. The “Generate t0 wave” procedure generates an empirical fit to the t0 Fit Coefficients using an appropriate function.



Choosing the function, TA Analysis → Generate t0 wave from Fit Coeffs, brings up a simple dialog:

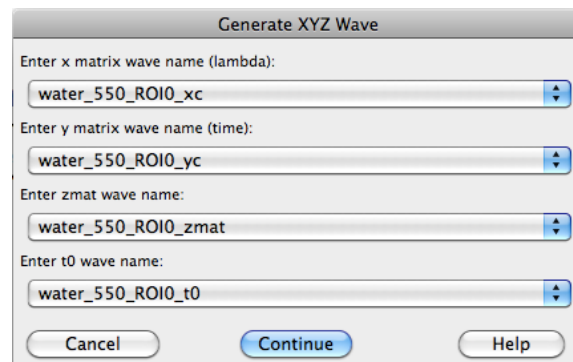
Enter the appropriate wave names as is described. Clicking “Continue” will generate two waves, one with a set of data points generated by the Fit Function which are used for plotting and evaluating the success of the fitting procedure (fit\_t0poly). The second is named *filename\_t0*. This is a 1D wave consisting of t0 values for each lambda point contained in the corresponding data file for a given set of  $\Delta A$  data.



This process should be repeated for each new data set that needs to be chirp corrected.

### Chirp Correction from t0 Wave

The final chirp correction function, TA Analysis → Chirp Correction from t0 Wave, can be called once you have generated *filename\_t0* for each data set that needs to be chirp corrected. The following files are generated:



Generate XYZ Wave

Enter x matrix wave name (lambda):  
water\_550\_ROI0\_xc

Enter y matrix wave name (time):  
water\_550\_ROI0\_yc

Enter zmat wave name:  
water\_550\_ROI0\_zmat

Enter t0 wave name:  
water\_550\_ROI0\_t0

Buttons: Cancel, Continue, Help

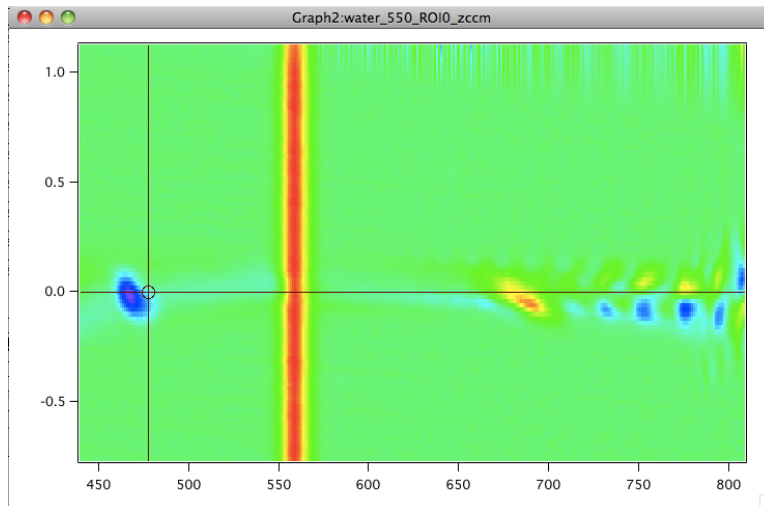
- *filename\_ccp* (parametric wave with non-interpolated z data and chirp corrected time values)
- *filename\_xcc* (1D wavelength data)

- *filename\_ycc* (chirp corrected 1D time data from first lambda value)
- *filename\_zccm* (z matrix of interpolated  $\Delta A$  data)
- *filename\_xcim* (wavelength values for image plot)
- *filename\_ycim* (corrected time values for image plot)

NOTE: The chirp correction process necessarily generates interpolated DA data for all wavelength points after the first. Kinetic fitting should be accomplished using non-interpolated data. This files should be used for visualization of spectra with the understanding that the data has been processed.

The parametric wave, *filename\_ccp* is not interpolated and can be visualized with Gizmo in 3D plots.

A chirp corrected image file will be automatically generated from *filename\_zccm* along with *filename\_xicc* and *filename\_yicc*. The standard analysis, extraction, and visualization tools can be used with this image.



## VIII. Data Analysis Using Singular Value Decomposition

### Introduction

Singular value decomposition is an analysis method that allows you to identify the number of linearly independent components within a given data set. When applied to time resolved absorption data, it is possible to obtain the number of distinct kinetic species and their associated wavelength dependent absorption coefficients. Furthermore, the SVD routines can be used to significantly improve signal/noise and to remove unwanted spurious features from a given data set. It can be used, among other things, for accurate determination of the characteristic lifetimes contained within a given data set by global fitting the results of the SVD.

These matrix analysis methods are native to the Igor Pro programming environment. These set of procedures provided are meant to provide a quick, User-friendly, and consistent interface to some common Igor functions. In this section, I will review some common uses of SVD for analysis of transient absorption data.

Additional reading on the subject:

*Singular Value Decomposition: Application to Analysis of Experimental Data.* E. R. Henry and J. Hofrichter, *Methods in Enzymology* 210 129 (1992).

*Visualization of transient absorption dynamics – towards a qualitative view of complex reaction kinetics.* H. Satzger and W. Zinth, *Chemical Physics* 295 287 (1993).

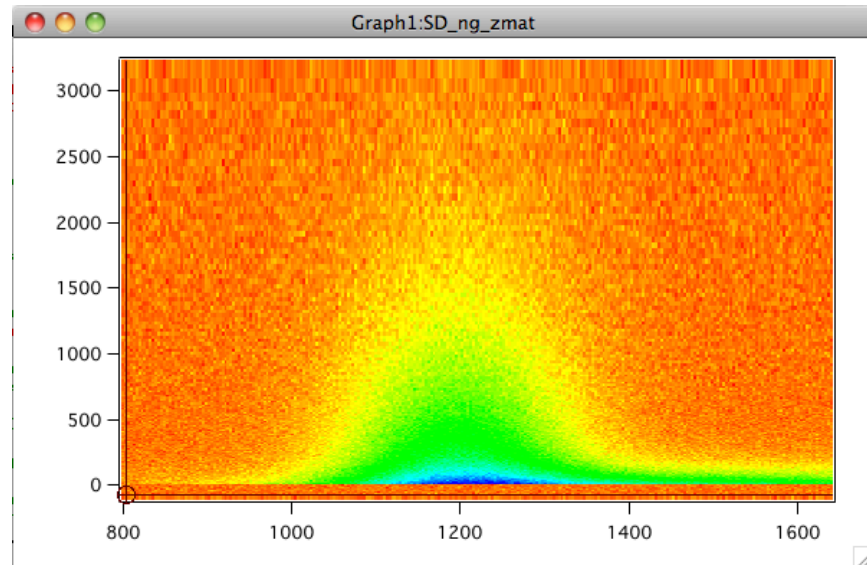
### Singular Value Decomposition

Briefly, singular value decomposition takes your data matrix and decomposes it into 3 matrices such that  $\Delta A = USV^T$ . If  $\Delta A$  is (n x m), then the resulting U is (n x n), S is (n X m), and V is (m x m). The S matrix is a diagonal matrix whose elements are the singular values. In the Igor implementation, S is a column vector of length n composed of the singular values.

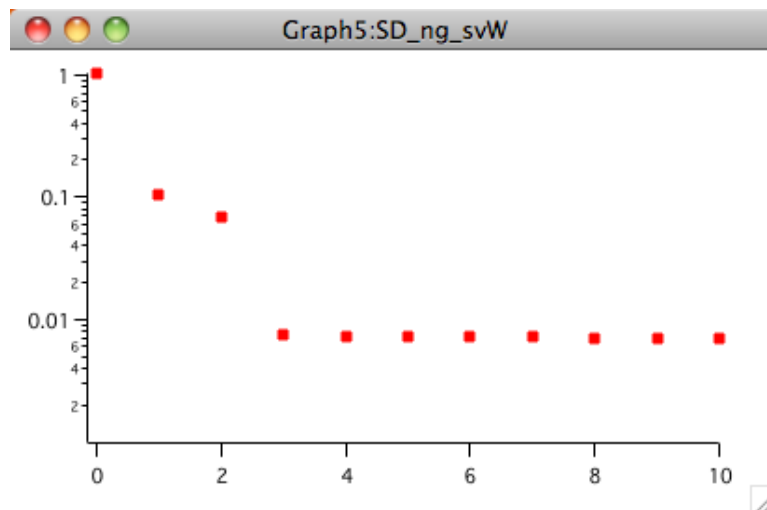
The matrices U and V are mathematical constructs. Without further processing, they have no physical meaning. However, they do represent linear combinations of the different spectral (U) and kinetic (V) components of the data set.

One major goal is to determine the rank of the matrix, in other words, the number of species or factors present in the experimental data. This can be determined by the number of non-zero singular values obtained in the decomposition. However, the presence of noise in the measurement can often make an unambiguous determination of the rank challenging. To assist in the analysis, the SVD procedure plots the singular values on a log scale. The challenge is to identify the cutoff, i.e., the noise level.

Here, we use an artificial data set with a set of 3 decaying Gaussian absorption profiles to illustrate the behavior of the procedures. We have added some Gaussian noise to the data set simulate the magnitude of the real experimental noise.



- 1) Click on the image file to make it the top graph. If necessary, extract a clean region of interest for the SVD analysis. Choose:  
TA Analysis → Singular Value Decomposition
- 2) The procedure will generate the SVD matrices and plot the first 10 singular values on a log scale for inspection:  
*filename\_psU* (U matrix from SVD)  
*filename\_svW* (list of singular values)  
*filename\_pkV* (V matrix from SVD)



## Rank SVD Matrices

- 1) It is now necessary to determine the rank of the data set by choosing the appropriate number of non-zero singular values. From the above data set, it is obvious that the correct number is 3 before the noise floor. However, for real data this is often a significant challenge. Choose:

TA Analysis → Rank SVD Matrices

- 2) The following files are generated:

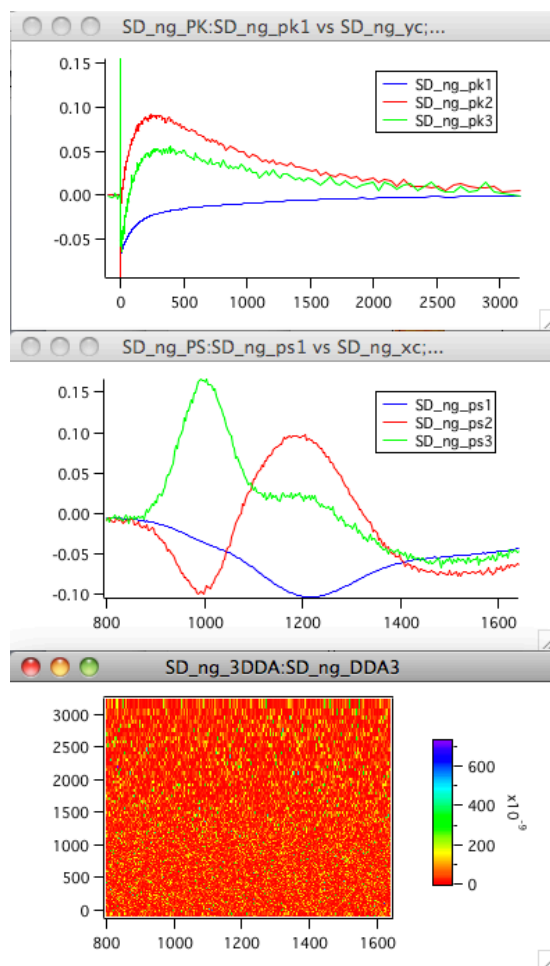
*filename\_ps*[1 ... *r*] (1D principal spectra)

*filename\_pk*[1 ... *r*] (1D principal kinetics)

*filename\_Arsv* (reconstructed data matrix from ranked SVD)

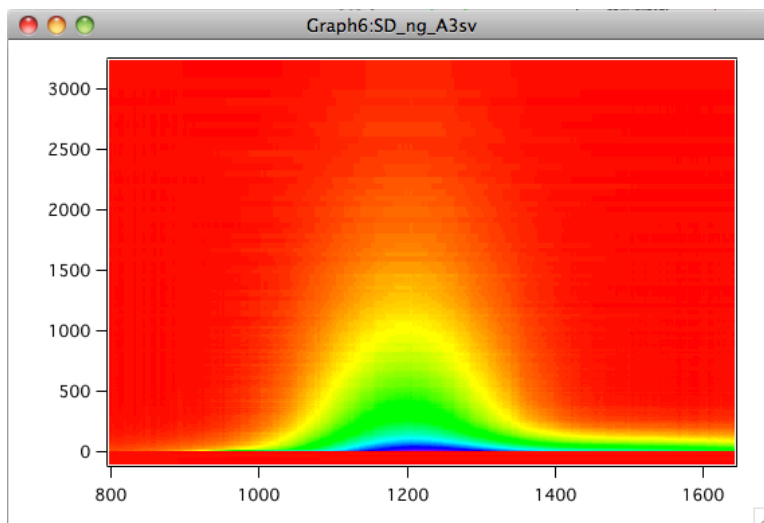
*filename\_DDAr* ( $\Delta\Delta A^2 = (\Delta A \text{ Data Matrix} - \textit{filename\_Arsv})^2$ )

- 3) By default, only an image plot of *filename\_DDAr* is automatically generated along with plots of the first *r* columns of the U and V matrices. You can choose to scale them by the square root of the respective singular value for visualization and analysis purposes. However, for fitting, you will need to make sure you generate unscaled principal spectra and kinetics.





- 4) Although the reconstructed matrix is not plotted by default, it is often useful for noise reduction. You can generate the image plot manually and then use the appropriate extraction procedures for presentation and analysis.



### Global Fit

This menu item provides a convenient shortcut to Igor's native Global Fitting procedure. It can also be found in the menu:

Analysis → Global Fit

It required the Global Fit 2.ipf procedure which can be found in the Igor Pro Folder under:

Igor Pro Folder → WaveMetrics Procedures → Analysis → Global Fit 2.ipf

It can be used to fit the individual principal kinetic files produced by the Singular Value Decomposition procedures or to fit multiple extracted spectra. Please refer to the Igor Pro help files for details and instructions.

### Species Absorption from Global Fit

This menu item is under development. This section will be updated in the next release.

## IX. Export a Progressive Time Axis for Data Acquisition

This procedure generates a set of measurement times in a 1D row format for use with common TA acquisition software. It uses an exponentially increasing spacing, similar to Megerle et. al., *Applied Physics B* 96 215 (2009).

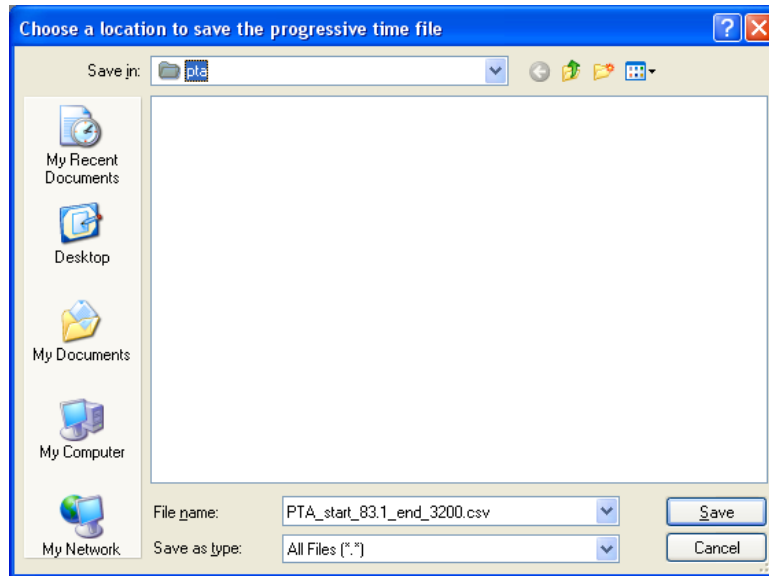
- 1) Choose the procedure from the TA Analysis Menu:  
TA Analysis -> Generate Progressive Time Axis
- 2) Enter the value of absolute time zero location in the dialog box. This value is referenced from the position of smallest delay on you mechanical stage, in units of ps.
- 3) Enter the value of the step size for the time window from -1 ps to +1 ps (relative delay to  $t_0$ ). In this range the step size is constant. From 1 ps until the end of the scan (entered into dialog box in relative ps) the number of points is reproduced over each order of magnitude in time (1 - 10 ps, 10 - 100 ps, etc.). The sparse pre-points option will insert 10 data points in the 100 ps before time zero if desired for display and baseline determination.



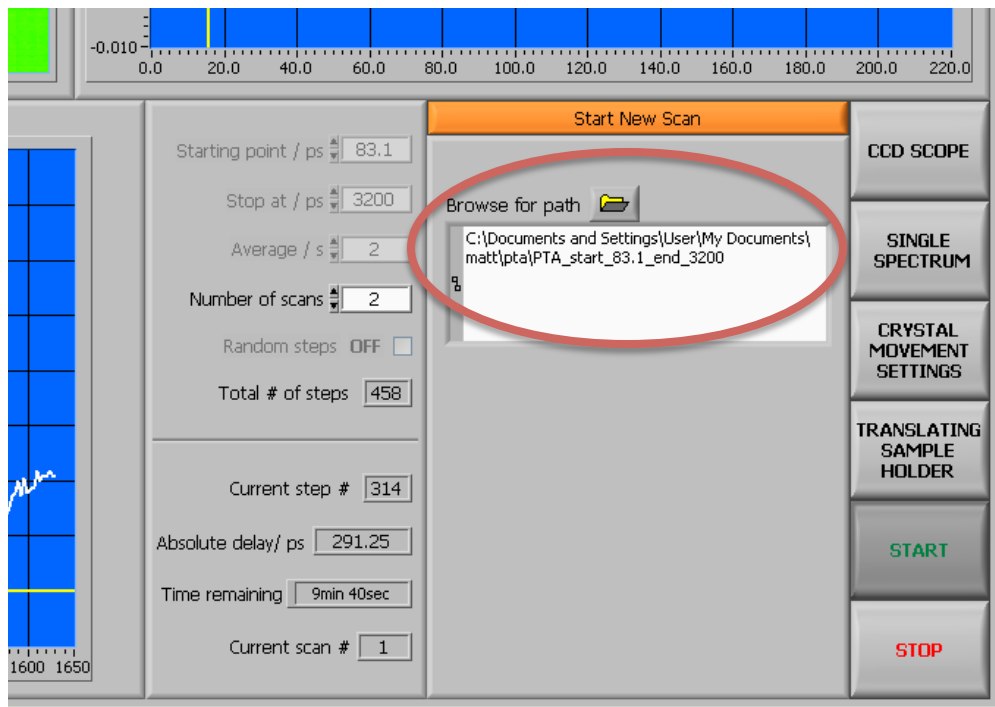
The screenshot shows a Windows-style dialog box titled "Generate Progressive Time Axis". It has a blue header bar with a question mark icon and a close button (X). The dialog contains the following elements:

- Label: "Enter time zero value (absolute ps):" followed by a text input field containing "184.1".
- Label: "Enter t0 step size (ps):" followed by a text input field containing "0.02".
- Label: "Enter max delay time (relative ps):" followed by a text input field containing "3200".
- Label: "Use sparse pre-points?" followed by a dropdown menu currently showing "yes".
- At the bottom, there are three buttons: "Cancel", "Continue", and "Help".

- 4) The procedure will automatically generate a dialog box so that the file can be saved in the correct format (.csv). It will automatically be given a name that defines the start and stop points (absolute and relative time respectively) based on the input parameters.



- 5) Load the file into the appropriate data acquisition software. The name of the file will give you the appropriate start and end points. If sparse pre-points are used, this will be 101 ps before time zero. Otherwise, it will be 1 ps.



## Appendix A: Automatically Generated Wave Names

For an imported data file, the first 22 characters of the filename will form the base for all automatically generated waves (herein referred to as *filename*).

### Load TA Data:

*filename\_zmat* (corrected matrix)  
*filename\_xc* (processed row values [wavelength])  
*filename\_yc* (processed column values [time])  
*filename\_xim* (x values for image plot [wavelength])  
*filename\_yim* (y values for image plot [time])  
*filename\_sca* (xyz wave for scatter plot)  
*filename\_par* (3D parametric wave)

### Extract Kinetics:

*filename\_kwav\_n* (kinetic data at wavelength *wav* averaged over *n* pixels)

### Extract Spectra:

*filename\_ktim\_n* (spectrum at time *tim* averaged over *n* pixels)

### Scattered Light Correction:

*filename\_scab* (averaged background spectrum)  
*filename\_zibg* (corrected data matrix)

### Extract Region of Interest:

*filename\_zroi* (subset of *zmat*)  
*filename\_xroi* (subset of *xc*)  
*filename\_yroi* (subset of *yc*)  
*filename\_ximr* (new image x coordinates)  
*filename\_yimr* (new image y coordinates)

### Import SX Pro Solvent Fit Coeffs

*filename\_sxfcn* (2D matrix of lambda, time zero pairs)

### Generate Fit Coeffs from Solvent Response

*filename\_kwav\_n* (kinetic data at wavelength *wav* averaged over *n* pixels)

### Generate t0 wave from Fit Coeffs

*fit\_t0poly* (result of order 3 polynomial fit to Fit Coeffs)  
*filename\_t0* (1D wave of t0 values for each lambda point)

**Chirp Correction from t0 Wave**

*filename\_ccp* (parametric wave with non-interpolated z data and chirp corrected time values)

*filename\_xcc* (1D wavelength data)

*filename\_ycc* (chirp corrected 1D time data from first lambda value)

*filename\_zccm* (z matrix of interpolated  $\Delta A$  data)

*filename\_xcim* (wavelength values for image plot)

*filename\_ycim* (corrected time values for image plot)

**Singular Value Decomposition:**

*filename\_psU* (U matrix from SVD)

*filename\_svW* (list of singular values)

*filename\_pkV* (V matrix from SVD)

**Rank SVD Matrices:**

*filename\_ps*[1 ... *r*] (1D principal spectra)

*filename\_pk*[1 ... *r*] (1D principal kinetics)

*filename\_Arsv* (reconstructed data matrix from ranked SVD)

*filename\_DDAr* ( $\Delta\Delta A^2 = (\Delta A \text{ Data Matrix} - \textit{filename\_Arsv})^2$ )

**Generate Progressive Time Axis:**

PTA\_start\_pos1\_end\_pos2 (time spacing between positions *pos1* and *pos2*)

*Note: The following files are generated by the Image Inspector and should be ignored:*  
kineticsA ; kineticsB ; spectraA; spectraB

## **Appendix B: Description of Fitting Functions and Coefficients**

This section outlines the structure and parameters of the supplied fitting functions.

Under development.